

# Stereoselective synthesis of belactosin C and its derivatives using a catalytic proline catalyzed crossed-aldol reaction<sup>☆</sup>

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**Abstract**—A highly practical and concise stereoselective total synthesis of belactosin C and synthetic variants was achieved using an *S*-proline catalyzed crossed-aldol reaction as the key step.

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Many natural products possessing the 2-oxetanone ( $\beta$ -lactone) moiety exhibit biological activity such as anti-biotic, antitumour and antiobesity.<sup>1</sup> Prominent natural molecules bearing a 2-oxetanone ring include anisatin, a potent vegetal poison and the antibiotic 1233A.<sup>2</sup> A significant number of natural 2-oxetanones with interesting biological activity have been identified and synthesized.<sup>3</sup> Recent additions to this family are belactosins A and C (Fig. 1) which inhibit the 20s proteasome *in vitro* ( $IC_{50} = 0.4 \mu\text{M}$ , chymotrypsin-like activity) in a yeast based assay of Streptomycin metabolites.<sup>4</sup>

The basic structure of the belactosins consists of a *trans*- $\beta$ -lactone moiety attached to an ornithine–alanine amino acid dipeptide unit. Degradation studies suggested that the  $\beta$ -lactone moiety is responsible for the antiproliferative activity. Two total syntheses of belactosins A and C and their analogues have been previously reported.<sup>5</sup> Recently, we developed a stereocontrolled route to belactosin C via an Oppolzer aldol reaction as the key step using a sultam chiral auxiliary.<sup>6</sup> Despite these synthetic methods, there is still a need for an alternative efficient synthetic route for belactosin C and new derivatives in order to further investigate  $\beta$ -lactones and their efficacy towards cancer and other diseases (Fig. 1). We envisioned that belactosin C could be read-

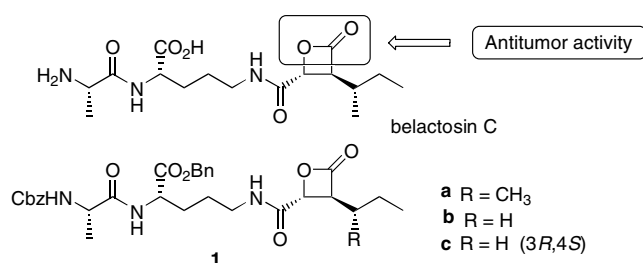


Figure 1.

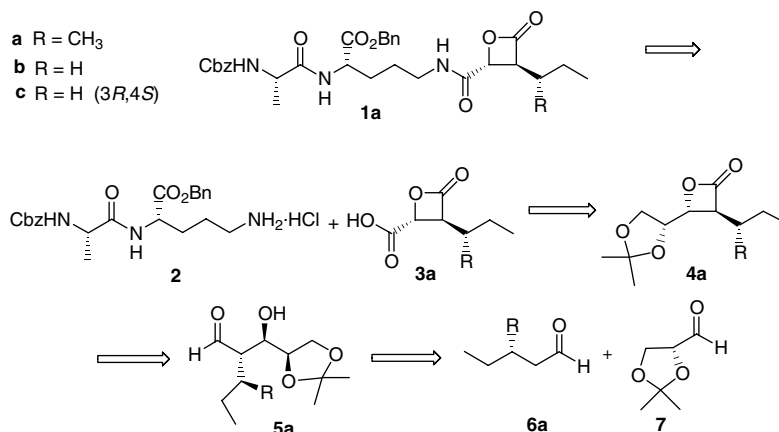
ily accessed by an *S*-proline catalyzed crossed-aldol reaction to install the two stereocentres of the *trans*- $\beta$ -lactone in a highly diastereo and enantioselective manner.<sup>7</sup> The retrosynthetic approach to belactosin C and derivatives are shown in Scheme 1.

In this Letter we report a concise and a highly stereoselective synthetic route to belactosin C wherein the chirality is derived from catalytic *S*-proline. Our synthesis began with a 10 mol % *S*-proline catalyzed crossed-aldol reaction between 3-(*S*)-methylvaleraldehyde **6a**, derived from *L*-isoleucine **8** and glyceraldehyde acetonide **7**,<sup>8</sup> which in turn was prepared from *D*-mannitol. After initial trials, the reaction was carried out with 10 mol % of *S*-proline and 1 equiv of **6a** and 2 equiv of **7** in dry DMF at 4 °C, stirring for 48 h led to aldol product **5a**. Due to the capricious nature of **5a**,<sup>9</sup> it was subjected to oxidation using sodium chlorite and sodium dihydrogen orthophosphate. The resulting  $\beta$ -hydroxy acid was lactonized with bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BOPCl) to give  $\beta$ -lactone acetonide **4a** in 46% overall yield from **6a**.

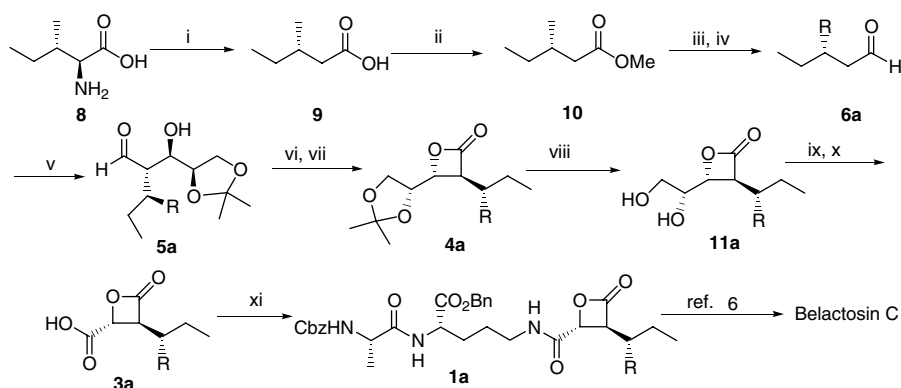
**Keywords:** *S*-Proline; Belactosin C;  $\beta$ -Lactone; Ornithine–alanine; Proliferative activity.

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Scheme 1.



**Scheme 2.** Reagents and conditions: (i) NaNO<sub>2</sub>, HBr, 0 °C–rt, 12 h; Zn, H<sub>2</sub>SO<sub>4</sub>, 0 °C–rt, 12 h (65% yield); (ii) dry MeOH, H<sub>2</sub>SO<sub>4</sub>, reflux, 12 h (70% yield); (iii) LiAlH<sub>4</sub>, Et<sub>2</sub>O, rt, 3 h; (iv) PCC, DCM, rt, 1 h (59% yield over two steps); (v) *S*-proline (10 mol %), DMF, addition of **6a** via syringe pump over 24 h; then 48 h, 4 °C; (vi) NaClO<sub>2</sub>, 20% NaH<sub>2</sub>PO<sub>4</sub>·2H<sub>2</sub>O, *t*-BuOH, 0 °C–rt, 4 h; (vii) BOPCl, Et<sub>3</sub>N, DCM, 23 °C, 1 h (46% overall yield from **6a**); (viii) 1 N HCl:THF (1:1) 0 °C–rt, 3 h (95% yield); (ix) NaIO<sub>4</sub>, 1,4-dioxane:H<sub>2</sub>O (1:2), 20 °C, 3 h; (x) NaClO<sub>2</sub>, 20% NaH<sub>2</sub>PO<sub>4</sub>·2H<sub>2</sub>O, *t*-BuOH, 0 °C–rt, 4 h (83% yield over two steps); and (xi) **2**, DCC, HOBT, EtOAc:H<sub>2</sub>O (1:1) rt, 1 h (50% yield).

β-Lactone acetonide **4a** was subjected to hydrolysis with 1 N HCl:THF (1:1) at 0 °C to rt, for 4 h to give β-lactone diol **11a** in 95% yield. Diol **11a** was subjected to oxidative cleavage using sodium metaperiodate in 1,4-dioxane:H<sub>2</sub>O (1:2) to give a crude aldehyde which was further oxidized with chlorite/dihydrogen orthophosphate to afford β-lactone carboxylic acid **3a** in an 83% yield over the two steps.<sup>10</sup> β-Lactone carboxylic acid **3a** was coupled with dipeptide **2**<sup>11</sup> which resulted cleanly in fully protected belactosin C **1a** in an 18% overall yield from aldol **5a**, (Scheme 2).

Belactosin C derivative **1b** was synthesized using valeraldehyde **6b** (R = H) and glyceraldehyde acetonide **7** followed by the same sequence of steps as used in Scheme 2 in an overall yield of 20%. In the same way, diastereomer **1c**<sup>12</sup> was prepared using *R*-proline instead of *S*-proline in an overall yield of 19%. The analytical data of **1a** and **1b** were in full agreement with the reported data.<sup>6</sup>

In conclusion, we have developed a catalytic route for the total synthesis of belactosin C and derivatives. Significantly, the chirality of the β-lactone moiety was

installed with inexpensive reagents. Application of this novel methodology to synthesize various substituted β-lactones in order to study their activity profiles is under progress.

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### Supplementary data

Experimental procedures, spectral data and copies of spectra for compounds **1c**, **3c** and **4a–c** can be found in the Supplementary data. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2007.01.059.

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8. Aldehydes containing an  $\alpha$ -methine carbon (CHR<sub>3</sub>) adjacent to the carbonyl do not undergo homo-dimerization when exposed to the catalyst as the kinetic inaccessibility of the  $\alpha$ -methine proton and thermodynamic instability of the corresponding enamine effectively prohibit nucleophile formation.
9.  $\beta$ -Hydroxyaldehydes such as **5a** were reported to undergo oligomerization, elimination and other decomposition reactions. For recent documented examples, see: (a) Chemler, S. R.; Roush, W. R. *J. Org. Chem.* **2003**, *68*, 1319–1333; (b) Lautens, M.; Stammers, T. A. *Synthesis* **2002**, 1933–2012.
10. (a) Efforts to estimate the level of diastereoselectivity of **5a** were not successful. Presumably, the minor diastereomer formed during the S-proline catalyzed aldol reaction was removed in the subsequent purification of **4a**; (b) The diastereo and enantioselectivity of aldol product **5a** was determined on the basis of  $\beta$ -lactone carboxylic acids **3a** and **3b** reported in our previous work.<sup>6</sup> The optical rotation of compound **3c**, was similar in magnitude but opposite in sign to that of **3b** ( $[\alpha]_{\text{D}}^{25} +13.8$  ( $c$  0.05, CHCl<sub>3</sub>); lit.<sup>6</sup> ( $[\alpha]_{\text{D}}^{25} -10.5$  ( $c$  1, CHCl<sub>3</sub>)).
11. For the preparation of dipeptide unit **2**, see Ref. 6.
12. Selected data: Compound **1c**:  $[\alpha]_{\text{D}}^{25} -11.5$  ( $c$  0.0125, CHCl<sub>3</sub>); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.27–7.30 (m, 10H), 5.67 (d,  $J = 8.0$  Hz, NH), 5.12 (d,  $J = 8.0$  Hz, 2H), 5.03 (s, 2H), 4.50–4.60 (m, 1H), 4.43–4.49 (m, 1H), 4.42 (d,  $J = 4.4$  Hz, 1H), 3.40–3.50 (m, 1H), 3.20–3.29 (m, 1H), 2.95–3.08 (m, 1H), 1.66–1.82 (m, 4H), 1.36–1.41 (m, 4H), 1.35 (d,  $J = 6.8$  Hz, 3H), 0.94 (t,  $J = 7.3$  Hz, 3H); <sup>13</sup>C NMR (75 MHz)  $\delta$ : 172.3, 171.5, 169.5, 167.9, 156.0, 136.1, 135.0, 128.6, 128.5, 128.4, 128.3, 128.1, 127.9, 72.8, 67.3, 67.0, 57.4, 51.7, 51.1, 38.4, 29.9, 29.1, 25.1, 19.8, 18.3, 13.4; IR (KBr): 3316, 2933, 1825, 1731, 1652, 1534, 1447, 1238, 1178, 1072, 905, 738, 661. MS (LC):  $m/z$  590 [M+Na]<sup>+</sup>. Compound **4a**:  $[\alpha]_{\text{D}}^{25} -28.6$  ( $c$  0.025, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 4.23 (ddd,  $J = 2.2, 6.8, 9.0$  Hz, 1H), 4.19 (dd,  $J = 2.2, 3.7$  Hz, 1H), 4.08 (dd,  $J = 6.8, 8.3$  Hz, 1H), 3.87 (dd,  $J = 6.8, 8.3$  Hz, 1H), 3.49 (dd,  $J = 3.7, 8.3$  Hz, 1H), 1.85–1.95 (m, 1H), 1.58–1.70 (m, 1H), 1.41 (s, 3H), 1.37 (s, 3H), 1.23–1.33 (m, 1H), 1.00 (d,  $J = 6.8$  Hz, 3H), 0.95 (t,  $J = 7.5$  Hz, 3H); <sup>13</sup>C NMR (75 MHz)  $\delta$ : 170.0, 110.2, 74.8, 73.0, 65.0, 57.9, 33.3, 26.7, 25.7, 25.4, 16.5, 10.9; IR (KBr): 2962, 2875, 1828, 1460, 1378, 1214, 1114, 1066, 895; MS (LC):  $m/z$  251 [M+Na]<sup>+</sup>. Compound **11a**:  $[\alpha]_{\text{D}}^{25} -38.5$  ( $c$  0.05, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 4.33 (dd,  $J = 3.9, 8.1$  Hz, 1H), 3.68–3.80 (m, 3H), 3.53 (dd,  $J = 4.3, 8.3$  Hz, 1H), 2.94 (br s, 1H), 2.30 (br s, 1H), 1.89–1.99 (m, 1H), 1.60–1.70 (m, 1H), 1.20–1.33 (m, 1H), 1.05 (d,  $J = 6.6$  Hz, 3H), 0.96 (t,  $J = 7.4$  Hz, 3H); IR (KBr): 3382, 2963, 1813, 1459, 1293, 1114, 1014, 877. MS (LC):  $m/z$  211 [M+Na]<sup>+</sup>.